

Product: **SFIM60 (Serum Free Insect Media)**
Solution
Catalog #: 900-630

Product Description

SFIM60 is a serum-free medium designed and optimized for the culture of Lepidopteran spp. insect cells. (Class, Insecta; Order, Lepidoptera; Family Noctuidae). The medium supports the growth and maintenance of both anchorage-dependent and suspension cultures of Sf-9 cells derived from the pupal ovarian tissue of the Fall Armyworm, *Spodoptera frugiperda* (J.E.Smith). The Sf-9 cell line is commonly used to isolate and propagate recombinant Baculoviral stocks and for the production of recombinant proteins. SFIM60 is also a protein-free medium that provides excellent results in comparison to such media as TNM-FH (Supplemented Grace's) in which 10% Fetal Bovine Serum is added. Last but not least, SFIM60 has shown excellent performance in the cultivation of high-V cells (i.e. *Trichoplusia ni* embryo cells) and the production of recombinant betagalactosidase.

Shelf life and Storage

SFIM60 should be stored under defined conditions between 2-8°C. The product should not be left in the light for prolonged periods as it is light-sensitive. When stored in the dark under ideal conditions, the product is stable until the expiry date on the label.

Instructions for use

- 1) Take a bottle from the proper storage conditions between 2-8°C.
- 2) Allow to warm to room temperature prior to use.
- 3) Ensure that the cap of the bottle is tight.
- 4) Gently swirl the solution in the bottle.
- 5) Wipe the outside of the bottle with a disinfectant solution such as 70% ethanol.
- 6) Using aseptic/sterile technique under a laminar-flow culture hood, work according to established protocols.

Serum-Supplemented Cultures:

Insect cells are incubated at 27-28°C in an open environment. SF-9 cells are usually cultured in Grace's medium supplemented with TPB and yeastolate (TNM-FH medium) plus 10% heat-inactivated FBS. Cells may also be cultured in IPL-41 medium containing 10% heat-inactivated FBS. A stationary culture will reach confluency in 3-6 days.

The cells are loosely attached to the substrate, and when the cells are subcultured, it is necessary to scrape the cells gently to remove them. To initiate a suspension culture, inoculate 3-5 x 10⁵ cells/ml from the stationary culture in a 250ml shaker flask, containing 100ml of complete medium. Subculture the cells to 3-5 x 10⁵ cells/ml twice weekly. The shaker should be maintained at 27-28°C. The caps of the flasks should be loosened for aeration.

Serum-Free Cultures:

Either weaning or direct adaption may be used for transferring cells from serum-containing media to SFIM60 Serum-Free Medium. We recommend using the weaning procedure for monolayer, as well as for suspension culture.

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Weaning Procedure:

1. Transfer cells in the logarithmic phase from the serum-containing medium into 50% (v/v) mixture of serum-supplemented medium and SFIM60.
2. Subculture the cells after 3 days and reduce the percentage of the serum-supplemented medium to 40%.
3. Continue with the subculturing of the cells every 3 days and with each passage reduce the concentration of the serum-supplemented medium by a further 10%.
4. On the sixth passage, the cells will be fully adapted to SFIM60 serum-free medium.

Maintenance of SF-9 cells in SFIM60 serum-free medium:

	Stationary culture	Suspension culture
Inoculation density	6-10 x 10 ⁴ cells/cm ²	1.5 x 10 ⁶ cells/ml
	2-3 times/week	Every 3-4 days
Subculture	Subculture the cells when the viable cell count reaches 40-50 x 10 ⁴ /cm ² , with greater than 90% viability	Subculture the cells when the viable cell count reaches 3-5 x 10 ⁶ /ml, with greater than 95% viability. After 5 days in culture, the cell density reaches 6-8 10 ⁶ cells/ml.

For *in vitro* laboratory use or further manufacturing only. Not for human use.